

PCR LAB Experience Tutorial

This step-by-step tutorial will guide you through using [Primer-BLAST](#) and making sense of the outputs.



1. Open [Primer-BLAST](#). This is a tool developed for scientists to design and optimize primer sequences. One of the things that it does really well is to search the database of genome sequences looking for places where given primer sequences would bind.

We will use a set of primers that RockEDU frequently uses with students in our research laboratory at The Rockefeller University, where we have students scrape off some cheek cells with a toothpick (no blood, it doesn't even hurt) and then mix them with the PCR reagents including the set of primers below:

2. Enter the primers into the appropriate forward and reverse primer sequence boxes:
 FORWARD: TTAAGTCCACCATTAGCACC
 REVERSE: GAGGATGGTGGTCAAGGGAC

3. You will NOT enter a PCR Template (because you want to see *all* of the matches that the database can show you) and you will leave all of the standard parameters as-is. Thus, the top of the form should now look like this:



- Finally, scroll down toward the bottom of the form. For the Database, type “nr” to search all sequences. Make sure the Organism field is also empty to search broadly.

Primer Pair Specificity Checking Parameters Note: Parameter values that differ from the default are highlighted in yellow

Specificity check ☒ Enable search for primer pairs specific to the intended PCR template

Search mode Automatic

Database nr

Exclusion ☐ Exclude predicted Refseq transcripts (accession with XM, XR prefix) ☐ Exclude uncultured/environmental sample sequences

Organism
Enter an organism name (or organism group name such as enterobacteriaceae, rodents), taxonomy id or select from the suggestion list as you type.
[Add more organisms](#)

Entrez query (optional)

Primer specificity stringency
Primer must have at least 2 total mismatches to unintended targets, including at least 2 mismatches within the last 5 bps at the 3' end.
Ignore targets that have 6 or more mismatches to the primer.

Max target size 4000

Allow splice variants ☐ Allow primer to amplify mRNA splice variants (requires refseq mRNA sequence as PCR template input)

Get Primers ☐ Show results in a new window ☒ Use new graphic view

- Click Get Primers and wait for your results

- If everything runs smoothly, your results should appear as a list, starting like this...

NIH U.S. National Library of Medicine NCBI National Center for Biotechnology Information Sign in to NCBI

Primer-BLAST » JOB ID:mJJHlyI2JN4D4CHILIUf11aeF0V7JQ_4eg

Primer-BLAST Results

Input PCR template: none
Specificity of primers: Target templates were found in selected database: Nucleotide collection (nt)
Other reports: [Search Summary](#)

Detailed primer reports

Primer pair 1

	Sequence (5'→3')	Length	Tm	GC%	Self complementarity	Self 3' complementarity
Forward primer	TTAACTCCACCATAGCACC	20	55.04	45.00	4.00	0.00
Reverse primer	GAGGATGGTGTCAAGGGAC	20	59.75	60.00	3.00	2.00

Products on target templates

>MN334614.1 Homo sapiens isolate 11_H7a1(1113002256_S32) haplogroup H7a1 mitochondrion, complete genome

product length = 440
Forward primer 1 TTAACCTCCACCATAGCACC 20
Template 15970 15989
Reverse primer 1 GAGGATGGTGTCAAGGGAC 20
Template 16409 16390

>MN334613.1 Homo sapiens isolate 6_H7a1c(1113001788_S82) haplogroup H7a1c mitochondrion, complete genome

product length = 440
Forward primer 1 TTAACCTCCACCATAGCACC 20
Template 15970 15989
Reverse primer 1 GAGGATGGTGTCAAGGGAC 20
Template 16409 16390

>MN334612.1 Homo sapiens isolate 1_H7a1(1113000994_S32) haplogroup H7a1 mitochondrion, complete genome

product length = 440

- The area of interest is the list of “Products on target templates” which tells you all of the sequences in the database that would produce a “hit” or positive signal in the PCR assay due to these primers binding and amplifying the identified sequence of DNA. Let’s break down the information that you’re given:



>[MN334614.1](#) Homo sapiens isolate 11_H7a1(1113002256_S32) haplogroup H7a1 mitochondrion, complete genome

```
product length = 440
Forward primer  1      TTAAGTCCACCATAGCACC  20
Template        15970 ..... 15989

Reverse primer  1      GAGGATGGTGGTCAAGGGAC  20
Template        16409 ..... 16390
```

MN334614.1 is the unique code that refers to this sequence in the genome database. The remainder of that line is the name given to that sequence. You'll notice that the naming isn't always consistent but gives you a pretty good idea of the organism and relevant information about its collection. **In this sample name, where does it tell you this is from human mitochondrial DNA?**

Product length tells how long the piece of DNA is between the two primers. **How long is this PCR product?**

Then it shows the forward primer and below it the numbers of the positions of the corresponding bases in the identified sequence. The dots tell you that the sequence forms a perfect match, while any letters in that space would identify the specific mismatches. Then, same thing for the reverse primer. **Do these primers make a perfect match?**

8. Now you can scroll down to see the range of matches or use Ctrl+F to do a search for relevant sequences or organism names, or to give you a count of the number of sequences found. **How many human sequences matched with these primers?**

